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(54) Title: **SAFE ATTENUATED BOVINE VIRAL DIARRHEA VIRUSES FOR USE IN PREGNANT COWS**

(57) Abstract: This invention relates to the use of specifically attenuated live BVD (bovine viral diarrhea) viruses for the preparation of a vaccine for use in the prevention and/or treatment of BVDV infections in breeding stocks of cattle, pregnant cows and for fetal protection in pregnant cows.

***Safe attenuated bovine viral diarrhea viruses
for use in pregnant cows***

Field of the invention

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The present invention relates to the use of specifically attenuated live BVD (Bovine Viral Diarrhea) viruses for the preparation of a vaccine for use in the prevention and/or treatment of BVDV infections in breeding stocks of cattle, pregnant cows and for fetal protection in pregnant cows.

- 10 The invention also relates to a method of treatment and/or prevention of BVDV infections in the above named group of cattle.

Background of the invention

- 15 Bovine Viral Diarrhea Virus (BVDV) is the causative agent of BVD and mucosal disease in cattle (Baker, 1987; Moennig and Plagemann, 1992; Thiel et al., 1996). Fetal infection during pregnancy can result in the resorption of the fetus, abortions as well as birth of immunotolerant calves which are persistently infected with BVDV. These calves lack or have very low neutralizing antibody titers and are continuously
20 shedding high amounts of virus. Next to acute or persistently infected bulls these calves are the major source for virus spreading and are therefore of primary importance in the epidemiology of this disease. The major economical impact of BVD results from high abortion rates, stillbirths, fetal resorption, mummification, congenital malformations, and birth of weak and undersized calves. For a detailed review of the
25 pathogenesis it is hereby referred to the article of Moennig and Liess of 1995 in its entirety.

Two major antigenic groups of BVDV (type I and II) have been described (Becher et al. 1999) which display limited cross neutralizing antibody reactions (Ridpath et al. 1994).

- 30 BVDV and other pestiviruses share the ability to cross the placenta of pregnant host animals. Present attenuated live BVDV vaccines also cross the placenta of seronegative heifers and result in clinical symptoms of wild type BVDV infections (see Moennig and Liess, Orban et al. 1983; Liess et al. 1984).

Present vaccines for the prevention and treatment of BVDV infections still have drawbacks (Oirschot et al. 1999).

Killed vaccines (inactivated whole virus) or subunit vaccines (conventionally purified or heterologously expressed purified viral proteins) are most often inferior to live vaccines in their efficacy to produce a full protective immune response even in the presence of adjuvants.

Live BVDV vaccines, although attenuated, are most often associated with serious safety problems. As mentioned above, they cross the placenta of pregnant cows and lead to clinical manifestations in the fetus and/or the induction of persistently infected calves. Therefore, they cannot be applied to breeding herds that comprise pregnant cows. Pregnant cows must to be kept isolated from vaccinated cattle to protect fetuses and may not be vaccinated themselves. Furthermore, revertants of attenuated live BVDV pose a serious threat to cattle. For conventionally derived attenuated viruses wherein the attenuation is attained by conventional multiple passaging, the molecular origin as well as the genetic stability remains unknown and the outbreak of revertants is unpredictable.

Live vaccines with defined mutations as a basis for attenuation would allow to avoid the disadvantages of the present generation of attenuated vaccines. A further advantage of said attenuating mutations lies in their molecular uniqueness which allows for use as distinctive labels for an attenuated pestiviruses and to distinguish them from pestiviruses from the field.

Such live and specifically attenuated pestiviruses with a high potential for induction of immunity as well as a defined basis of attenuation which can also be distinguished from pathogenic pestiviruses are described in the PCT-application PCT/EP 99/03642. In this application it has been demonstrated that pestiviruses can be specifically attenuated by the inactivation of the RNase activity residing in glycoprotein E^{RNS}. The inventors disclose that this attenuation principle is universal to all pestiviruses and particularly suitable for BVDV.

This application discloses an attenuated live vaccine that is superior in its efficacy of induction of immunity to subunit and killed vaccines as well as superior to conventionally attenuated vaccines which are molecularly undefined, unmarked and unpredictable with respect to mutants. From prior art an expert would naturally associate the risk of fetal BVDV infection when vaccinating pregnant cows or cattle in

the near vicinity of pregnant cows with an attenuated live virus. This remains a serious drawback limiting the use of such a vaccine.

Therefore, the technical problem underlying this invention was to provide safely
5 attenuated and live BVDV vaccines that can be used for the preparation of a live vaccine for use in the prevention and/or treatment of BVDV infections of pregnant cows or cattle in the presence of pregnant cows.

Disclosure of the invention

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The solution to the above technical problem is achieved by the description and the embodiments characterized in the claims.

It has surprisingly been found that live BVDV (bovine viral diarrhea virus), wherein the RNase activity residing in its glycoprotein E^{RNS} is inactivated, does not result in
15 placental transmission to the fetus when pregnant cows are vaccinated with said viruses. No clinical signs in the calves are observable when a BVDV strain of said characteristic is used to challenge pregnant cows.

Therefore, in one aspect, the present invention relates the unexpected use of a live BVDV (bovine viral diarrhea virus), wherein the RNase activity residing in its
20 glycoprotein E^{RNS} is inactivated, for the preparation of a live vaccine for use in the prevention and/or treatment of BVDV infections in breeding stocks of cattle.

Even more unexpected, pregnant cows themselves may be vaccinated with said specifically attenuated BVDV. Consequently, in a preferred embodiment, the present invention relates to the use of said BVDV for the preparation of a live vaccine for use
25 in the prevention and/or treatment of BVDV infections in pregnant cows.

This allows for the first time ever to prevent and protect the fetus itself against BVDV infections with a live vaccine. In a more preferred embodiment, the present invention relates to the use of said BVDV for the preparation of a live vaccine for inducing fetal protection against BVDV infections in pregnant cows.

30 The term "vaccine" as used herein refers to a pharmaceutical composition comprising at least one immunologically active component that induces an immunological response in an animal and possibly but not necessarily one or more additional components that enhance the immunological activity of said active component. A vaccine may additionally comprise further components typical to

pharmaceutical compositions. The immunologically active component of a vaccine may comprise complete live organisms in either its original form or as attenuated organisms in a so called modified live vaccine (MLV) or organisms inactivated by appropriate methods in a so called killed vaccine (KV). In another form the immunologically active component of a vaccine may comprise appropriate elements of said organisms (subunit vaccines) whereby these elements are generated either by destroying the whole organism or the growth cultures of such organisms and subsequent purification steps yielding in the desired structure(s), or by synthetic processes induced by an appropriate manipulation of a suitable system like, but not restricted to bacteria, insects, mammalian or other species plus subsequent isolation and purification procedures or by induction of said synthetic processes in the animal needing a vaccine by direct incorporation of genetic material using suitable pharmaceutical compositions (polynucleotide vaccination). A vaccine may comprise one or simultaneously more than one of the elements described above.

Additional components to enhance the immune response are constituents commonly referred to as adjuvants, like e.g. aluminiumhydroxide, mineral or other oils or ancillary molecules added to the vaccine or generated by the body after the respective induction by such additional components, like but not restricted to interferons, interleukins or growth factors.

A "pharmaceutical composition" essentially consists of one or more ingredients capable of modifying physiological e.g. immunological functions of the organism it is administered to, or of organisms living in or on its surface like but not restricted to antibiotics or antiparasitics, as well as other constituents added to it in order to achieve certain other objectives like, but not limited to, processing traits, sterility, stability, feasibility to administer the composition via enteral or parenteral routes such as oral, intranasal, intravenous, intramuscular, subcutaneous, intradermal or other suitable route, tolerance after administration, controlled release properties.

A vaccine of the invention refers to a vaccine as defined above, wherein one immunologically active component is a live BVDV, wherein the RNase activity in its protein E^{RNS} is inactivated.

The term "live vaccine" refers to a vaccine comprising a living, in particular, a living viral active component.

The term "BVDV" as used herein refers to all viruses belonging to species BVDV 1 and BVDV 2 in the genus pestivirus, within the family Flaviviridae (Becher et al. 1999).

The more classical BVDV type I strains and the more recently recognized BVDV type II strains display some limited but distinctive differences in nucleotide and amino acid sequences. These differences are immunologically distinguishable by monoclonal antibodies. The attenuation principle of inactivating the RNase activity in the protein E^{RNS} as described in the above referenced international application is universally applicable and holds true for both type I and type II BVDV. The surprising feature of BVDV which are attenuated in said manner that these viruses do not cross the placental barrier and do not lead to clinical manifestations of BVDV infections in the fetus applies to all types of BVDV. Therefore, in a further more preferred embodiment the present invention relates to the use of said BVDV for the preparation of a live vaccine for the treatment and or prevention of fetal infection, pregnant cows and breeding stocks of cattle.

"RNase activity" as used herein refers to the ability of the glycoprotein E^{RNS} which is an inherent protein to all naturally occurring pestiviruses such as BVDV to hydrolyze RNA.

It should be noted that the term glycoprotein E0 is often used synonymously to glycoprotein E^{RNS} in publications.

The term "inactivation of the RNase activity residing in said glycoprotein" refers to the inability or reduced capability of a modified glycoprotein E^{RNS} to hydrolyze RNA as compared to the unmodified wild type of said glycoprotein E^{RNS}.

Inactivation of the RNase activity residing in glycoprotein E^{RNS} can be achieved by deletions and/or mutations of at least one amino acid of said glycoprotein as demonstrated in the international application PCT/EP99/03642 and by Hulst et al. (1998). Therefore, in a most preferred embodiment the present invention relates to the use of live BVDV, wherein said RNase activity is inactivated by deletions and/or mutations of at least one amino acid of said glycoprotein.

The amino acid sequence for the BVDV strain CP7 is accessible for reference purposes in the GenBank/EMBL data library (accession number U63479). Two regions of amino acids are highly conserved in glycoprotein E^{RNS} as well as in some plant and fungal RNase-active proteins (Schneider et al., 1993). These two regions are of particular importance to the RNase enzymatic activity. The first region consists

of the region at the amino acids at position 298 to 310 and the second region consists of the amino acids at position 341 to 360 of said viral polyprotein as exemplified for the CP7 strain of BVDV (numbering according to the deduced amino acid sequence of the CP7 strain; see Figure 1 for the amino acid sequence of the CP7 glycoprotein E^{RNS}). The amino acids of particular importance to the RNase activity as mentioned above are by no means limited to the exact position as defined for the CP7 strain of BVDV but are simply used in an exemplary manner to point out the preferred amino acids being at that position or corresponding to that position in BVDV strains since they are highly conserved. For different BVDV pestiviruses the numbering of the positions of the preferred amino acids might be different but an expert in the field of the molecular biology of pestiviruses will easily identify these preferred amino acids by their position relative to the highly conserved amino acids of said glycoprotein.

As a consequence, the present invention relates in a more preferred embodiment to the use of BVDV according to the invention, wherein said inactivating deletions and/or mutations are located at the amino acids at position 298 to 310 and/or position 341 to 360, as described for the BVDV CP7 strain in an exemplary manner and corresponding thereto in other BVDV strains, of said glycoprotein.

In a most preferred embodiment the present invention discloses the use of a live BVD virus according to the invention wherein said RNase activity is inactivated by the deletion of the histidine residue at the position 349 as described for the CP7 strain of BVDV in an exemplary manner or corresponding thereto in other BVDV strains, of said glycoprotein.

In a further aspect the invention relates to a method for the prevention and/or treatment of BVDV infections in breeding stocks of cattle characterized in that a pharmaceutical composition comprising a live BVD virus, wherein the RNase activity residing in its glycoprotein E^{RNS} is inactivated, is applied to an animal in need of such prophylaxis or treatment.

To practice the invention it is necessary to prepare a BVDV according to the invention. This can be done with any available BVDV that is suited for recombinant modification. One example how to obtain such a virus according to the invention is given below in example 2. The example demonstrates the preparation of a virus for a use according to the invention in an exemplary manner and is by no means limiting. An expert in the field of virology or molecular biology can achieve this by standard

methods which are abundantly provided in the art.

The application of the preferred live viruses according to the application to the preferred group of animals is identical to the application methods which are available and practiced for other live attenuated viruses in the art.

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20

Examples

Example 1 BVDV fetal challenge study

Experimental design

- 25 Six pregnant heifers were selected from a BVDV negative herd. Two groups of 3 heifers each were included in the trial. The animals in the groups were kept isolated, thereby insuring the absence of cross and outside infection.

30

Group No	No. of animals	Virus	Application route	Dose per route TCID ₅₀
1	3	NCP7	i.m.; i.n.; i.t.; s.c.;	$1,2 \times 10^5$
2	3	B-349-d	i.m.; i.n.; i.t.; s.c.;	$1,2 \times 10^5$

Table 1

Heifers from groups 1 to 2 were inoculated with at least 10^5 TCID₅₀ BVDV in a volume of 3 ml per route. Application routes were intramuscular (i.m.), subcutaneous (s.c.), intranasal (i.n.) (1.5 ml per nostril) and intratracheal (i.t.).

Blood samples were collected from the animals for serology and virological examinations. Heifers were regularly monitored for the presence of clinical signs of BVDV infection during the observation period. The experiment was terminated after the birth of calves by examination for BVDV infection. Main evaluation parameters were the number of BVDV-related abortions and the number of calves born viraemic or comprising antibodies to BVDV.

Buffy coat preparations

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Blood was collected in suitable sterile vessels pre-filled with heparin solution (Na-heparin for injection, 5 000 IU/ml:Gedeon Richter RT, Budapest, Hungary) assuring at least 20 IU heparin per ml blood in the blood sample.

20

Blood was centrifuged for 10 min at 750 g at 4 °C. Erythrocytes sedimented in the lower half of the tube while the plasma made up the supernatant in the upper half of the tube. On top of the erythrocyte phase, in the so called interface, the leukocytes were located. This thin layer of white blood cells was collected with a sterile pipette and then resuspended very carefully in a new tube filled up with Gey's buffer (140mM NH₄CL, 2,7mM KCl, 6,5mM Na₂HPO₄, 1,5mM KH₂PO₄). The suspension was kept on ice for 10 minutes. Cells were centrifuged again (10 min, 750 g, 4 °C) and the supernatant discarded. Gey's buffer treatment was repeated until no substantial amount of erythrocytes was observed in the suspension. Then the cells

were washed twice in RPMI 1640 (Gibco BRL, Eggenstein, Germany) with 5 % FCS (fetal calf serum, about 20 ml) and centrifuged thereafter. The obtained buffy coats were resuspended in a small volume (2 ml) of RPMI 1640 5 % FCS and frozen at -70 °C in two aliquots of 1 ml. Buffy coats were used for the determination of blood cell associated BVDV.

BVDV antibody ELISA-test

At least 10 ml fresh blood were collected at each sampling time point from each animal. Blood was allowed to clot at room temperature, and separated by centrifugation. Each serum sample was tested for the presence of BVDV-antibodies using a suitable and validated ELISA test (Svanovir™ BVDV antibody test Cat# 10 2200-10). The test was validated and performed according to the manufacturer's recommendations. Positive samples were diluted according to the log₂ scale to determine BVDV antibody titers.

Fetal infection challenge viruses

The strain B349-d was applied to group 2. It is almost identical to the NCP7 strain that was applied to group 1 except for one single deletion of the histidine in position 349 of its amino acid sequence. B349-d was prepared according to the procedure listed below.

Detection of BVDV from buffy coat or tissue samples by virus isolation procedure followed by an indirect Immune Fluorescence Test (IFT)

For virus isolation a 20% suspension of tissues, serum or buffy coat were used. 1 g organ material was homogenized using a mortar and pistill, diluted up to 5 ml phosphate buffered saline (PBS) and centrifuged at 1500 * g for 10 min at 4°C. The supernatant was filtrated sterilly using 0,45 µm filter holders, Schleicher and Schuell, Dassel, Germany, and used for virus isolation.

Virus propagation was performed on Marbin Darby Bovine Kidney cells (MDBK) as monolayer in 24 well plates with EMEM, containing 10% fetal calf serum, 1%

nonessential amino acids and 0,15 % antibiotics. Before starting the cells were tested for contamination with BVDV by RT-nPCR as described below, and by immunofluorescence assay (IFA).

After splitting, the cells were each infected simultaneously with 100 µl suspension, serum or buffy coat in duplicates. 5 days later, one aliquot of the cell cultures was frozen and thawed.

100µl of the cell lysate was passaged twice to MDBK cell suspensions. One aliquot was fixed with ethanol and stored at -20°C for virus detection by immune fluorescence assay(IFA).

- 10 Infected cell cultures were tested by IFA. Cells were fixed by incubation with cold ethanol at -20°C. Fixed 24 well plates were washed with PBS twice. Afterwards the wells were incubated for two hours at room temperature with αBVDV-mAb-mix (Weiland et al. 1989). The αBVD-mAb-mix was diluted 1:10 in PBS. After washing three times with PBS rab.α-mouse FITC- conjugated antiserum (Jackson
- 15 ImmunoResearch Laboratories Inc., West Grove, PA 19390, USA) was added in a solution of 1:150 in PBS (phosphate buffered saline) and incubated for 1 h at room temperature. After additional washing, glycerol-PBS (1:1) was added and stained cell cultures were investigated by immunofluorescence microscopy.

20 Detection of BVDV from serum, buffy coat and tissue samples by Polymerase chain reaction (PCR)

- RNA was extracted by column chromatography using QIAamp Viral Mini Kit for serum samples and RNeasy Mini Kit for organ material and buffy coat, according to
- 25 the manufacturers recommendations. Both kits are available from Qiagen LTD, Hilden, Germany.

RT-nPCR:

- 30 Primer pairs for the detection of pestiviruses by RT-nPCR were used as previously described by Sandvik et al. (1997). The sequences of the external primers are:
- (P103F) '5-TAG CCA TGC CCT TAG TAG GAC T-3'
- (P365R) '5- TGT GCC ATG TAC AGC AGA GAT T -3'

The external primers generated a DNA- fragment of 280-284 bp length.

The sequences of the internal primers were as follows:

(B145F) `5-AAC AGT CCG TGA TTC GTT GGA T-3`

(B314R) `5-CAC CCT ATC AGG CTG TAT TCG T-3`

5 The internal primers generated a DNA- fragment of 191 bp length.

A volume of 5 µl total RNA (1µg – 1pg) was used for RT-PCR amplification, using the Titan One Tube Kit, Boehringer Mannheim, Mannheim, Germany. RT-PCR was carried out in a total volume of 25µl, containing 1*RT-PCR, 0,1 mM dNTPs, 20 pmol of sense and antisense primer, 5 mM dithiothreitol, 1 mM magnesium chloride, 2,5 –
10 5 U RNasin, Promega Ltd, 1-2,5 U enzyme mixture, AMV-RT and Taq Polymerase, ad 25µl Aqua dest. including 0,1% diethylpyrocarbonat.

RT-PCR-cycle conditions were as follows:

42°C for 1 hour, 95°C for 5 min and 35 cycles of 94°C for 1 min, 60°C for 1 min and 72°C for 1 min, finally prolongation for 10 min.

15 1 µl PCR product was used as template for the nested PCR, using Qiagen Taq (Qiagen LTD, Hilden, Germany).

nPCR was carried out in a total volume of 50µl, containing 1*PCR buffer, 3,5mM magnesium chloride, 0,1 mM dNTPs, 20 pmol of each primers and 2,5 U Taq.

The temperature profile of the nested PCR for 30 cycles was 94°C for 1 min, 55°C
20 for 1 min and 72°C for 1 min, followed by a prolonged last amplification.

A volume of 5 µl nPCR product was analysed by horizontal agarose gel electrophoresis in tris-acetate EDTA buffer, using 1% agarose and a 1 µg/ml ethidium bromid staining. The amplificate was identified by size comparison with a standard molecular weight 100 bp ladder (Gibco BRL, Eggenstein, Germany), the
25 specificity of the PCR products was checked by sequence analysis.

To run non-crosscontaminating PCRs the RoboAmp 4200 (MWG Biotech, Ebersberg, Germany) was used for automated pipetting. Thermocycling was performed in a DNA Thermal Cycler Primus 96 (MWG Biotech, Ebersberg, Germany) or T3 (Biometra, Göttingen, Germany).

30

Results

Clinical observation

5 No BVD related clinical signs were observed in Group 2 heifers during the observation period.

In Group 1, mostly increased salivation and nasal discharge were noticed for about a period of 6 days (between PI days 7 – 12). Two animals had diarrhea on day 14 PI. Results are summarized in table 2.

10 Table 2

Days PI	Ranking of heifer No. in					
	Group 1			Group 2		
	11093	13178	11736	12031	10565	11060
1	-	-	-	-	-	-
2	-	-	-	-	-	-
3	-	-	-	-	-	-
4	-	-	-	-	-	-
5	-	-	-	-	-	-
6	-	-	-	-	-	-
7	3	2	2	-	-	-
8	3	2	2	-	-	-
9	3	2	2	-	-	-
10	2	2	2	-	-	-
11	1	1	1	-	-	-
12	1	1	1	-	-	-
13	-	-	-	-	-	-
14	2	-	1	-	-	-
15	-	-	-	-	-	-
16	-	-	-	-	-	-
17	-	-	-	-	-	-
18	-	-	-	-	-	-
19	-	-	-	-	-	-
20	-	-	-	-	-	-
21	-	-	-	-	-	-

Ranking: 1 = mild, 2 = moderate, 3 = severe

Pregnancy history

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In Group 2, all heifers had a normal length of pregnancy (in a range of 270 – 277 days) and a pregnancy and parturition with no complication.

In Group 1, one animal (#11736) had abortion in the 8th month of pregnancy. Necropsy of the fetus revealed an acute sero-fibrinous inflammation of the chorion that led to an insufficient oxygen supply of the fetus and then to its

20

suffocation The remaining two animals gave birth to a calf after 278 (#13178) and 296 (#11093) days of pregnancy, respectively.

Virus detection from clinical samples obtained from aborted material or new born calves

In case of an abortion samples from different organs such as brain, spleen or kidney were investigated by PCR and/or IFT for BVDV. All clinical samples obtained from the aborted fetus of cow 11736 in group 1 were BVDV negative. Buffy coat samples from calves born from cows 11093 and 13178 were BVDV positive as confirmed by IFT and PCR. In contrast, buffy coats and serum samples from calves born from cows 10565, 110606 and 12031 of group 2 were all negative fro BVDV as confirmed by the above mentioned methods. The data is summarized in table 3.

Table 3

Fetal Transmission Evaluation						
Group	Animal No.	chall. Virus	calf No.	abort/ life	conclusion from virus detection data	percentage placental transmission of group
1	11093	NCP7	11093 c	live	pos	66%
	11736		11736 c	aborted	neg	
	13178		13178 c	live	pos	
2	10565	B349-d	10565 c	live	neg	0%
	11060		11060 c	live	neg	
	12031		12031 c	live	neg	

Serological data from pregnant cattle

At selection and day of infection the pregnant cows were BVDV negative. Serum samples from all animals in the trial collected 1 month after infection were BVDV

seropositive. Serum positivity was confirmed for BVDV throughout the trial by reinvestigating serum samples obtained on a monthly basis from all animals. The data is summarized in table 4.

5 Table 4

ANTIBODY ELISA DATA

Cow No.	Group	BVDV specific antibodies in sera taken at									
		Sele- tion	Infec- tion	PI month						At termination	
				1	2	3	4	5	6	cow	calf*
11093	1	-	-	+	+	+	+	+	+	+	-
13178	1	-	-	+	+	+	+	+	+	+	-
11736	1	-	-	+	+	+	+	Ab. [°]	NA	NA	NA
11060	2	-	-	+	+	+	+	+	+	+	-
10565	2	-	-	+	+	+	+	+	+	+	-
12031	2	-	-	+	+	+	+	+	-	-	-

PI = Post infection; * before the first ingestion of colostrum;

NA = Not applicable;

° after abortion, cows were removed from the study, Ab.=abortion

Discussion and conclusion

10 As 2 of 3 calves born in group 1 which were infected with NCP7 were viraemic for BVDV at birth whereas no calf born from group 2 which had been infected with B-349-d this data provides clear evidence that the genomic modification introduced in B-349-d abrogates the ability of the parent virus NCP7 to induce fetal infection.

15 Example 2 Generation of RNase-negative BVDV mutants

Starting with the full length cDNA clones pA/BVDV (Meyers et al., 1996b), from which infectious cRNA can be obtained by in vitro transcription, subclones were generated. A XhoI/BglII fragment from pA/BVDV was cloned into plasmid pCITE-2C,

cut with the same enzymes. Single stranded plasmid DNA was produced from these constructs according to the method of Kunkel (Kunkel et al., 1987) using E. coli CJ 236 cells (BioRad) and the VCMS single strand phage (Stratagene). The single stranded DNA was converted to double strands using the 'Phagemid in vitro Mutagenesis Kit' (BioRad). A synthetic oligonucleotide which was used as a primer for generating the desired BVDV mutant is listed below in an exemplary fashion:

B-349-d: CATGAATGGAACAAAGGTTGGTGCAACTGG

- 10 The double stranded plasmid DNA was used for transformation of E.coli XL1-Blue cells (Stratagene). Bacterial colonies harboring plasmids were isolated via ampicillin selection. Plasmid DNA was prepared and further analyzed by nucleotide sequencing using the T7 polymerase sequencing kit (Pharmacia). Plasmids containing the desired mutations and no second site changes were used for the
- 15 construction of full length cDNA clones. To obtain the BVDV CP7 mutant, a XhoI/BglII fragment containing the deletion was inserted into pA/BVDV cut with XhoI and NcoI together with a BglII/NcoI fragment isolated from pA/BVDV/Ins-. From construct pA/BVDV/Ins- a cRNA was transcribed that gives rise to a noncytopathogenic BVDV upon transfection in suitable cells (Meyers et al., 1996b).
- 20 The different full length clones were amplified, and the plasmids isolated. The presence of the desired mutations was proven by DNA sequencing. After linearization with SmaI cRNA was transcribed as described previously (Meyers et al., 1996ab). RNA was purified by gel filtration and phenol/chloroform extraction and used for transfection of bovine kidney (MDBK clone B2) cells. The transfections were
- 25 analyzed by immunofluorescence with virus specific antisera. The desired mutant could be recovered (as confirmed by immunofluorescence). The virus was amplified by passage on the same cell line used for the transfection experiments.
- Further analysis of the mutant included determination of one step growth curves and characterization of viral RNA by Northern blot with virus specific cDNA probes as well
- 30 as reverse transcription polymerase chain reaction (RT-PCR) and subsequent sequencing of the PCR fragments to verify the presence of the desired mutations in the viral genome. The recovered viruses grew equally well and produced similar amounts of RNA just as the virus resulting from the plasmid displaying the wild type sequence.

- The viability of the BVDV mutant was shown by transfection of the respective cRNA and splitting of the cells 3 days thereafter. Part of the cells was seeded into a 3.5 cm diameter dish, fixed with acetone/methanol at the day thereafter and analyzed by immunofluorescence with a mixture of BVDV-specific monoclonal antibodies (Weiland et al., 1989). All cells were found positive whereas a control of cells transfected with noninfectious RNA showed no signal. From a part of the cells transfected with the respective cRNA, an extract was produced by one cycle of freezing and thawing. Fresh cells were infected with this cell extract and proved to be BVDV positive by BVDV specific immunofluorescence 3 days post infection.
- Table 1 presents the changes introduced into the conserved sequences of E^{RNS} representing the putative active site of the RNase which are encoded by the virus mutant.

Table 1

Name	Sequence in RNase motif	RNase activity	Viability of mutant
pA/BVDV	...SLHGIWPEKIC...RHEWNKHGWCNW..	+	+
B-349-d	...SLHGIWPEKIC...RHEWNK_GWCNW..	-	+

- Legend to Table 1: Test for RNase activity was done in a transient assay. BHK21 cells were infected with Vaccina virus vTF7-3 (Fuerst et al, 1986) and then transfected with the respective cDNA construct (5 µg of plasmid DNA, transfection using Superfect as recommended by the supplier (Qiagen)). After 10 hours incubation at 37 °C in a CO₂ incubator, the transfected cells were lysed and processed for determination of RNase activity as described below). Viability was determined as described below.

Example 2 Effect of the mutation in position 349 of the BVDV strain CP7 on the RNase activity of E^{RNS}

- To test the effect of the deletion in position 349 on the RNase activity of E^{RNS} appropriate cells were infected with the mutant viruses.
- For the BVDV mutant RNase activity was tested with material obtained after RNA transfection without passage of the recovered viruses. Cells transfected with the appropriate RNA were split 72h post transfection and seeded in two dishes. 24h

later, from one dish, cell extracts were prepared and analyzed for RNase activity. Infection with wild type virus served as a positive control whereas noninfected cells were used as a negative control. Cells were washed twice with phosphate buffered saline and lysed in 0.4 ml of lysis buffer (20 mM Tris/HCl; 100 mM NaCl, 1 mM EDTA, 2 mg/ml bovine serum albumin; 1% Triton X100; 0.1% deoxycholic acid; 0.1% sodium dodecyl sulfate). The lysate was given into 1.5 ml reaction tubes, sonified (Branson sonifier B12, 120 Watt, 20 s in a cup horn water bath), cleared by centrifugation (5 min, 14,000 rpm, Eppendorf Centrifuge, 4°C) and the supernatant subjected to ultracentrifugation (Beckmann table top ultracentrifuge, 60 min at 4°C and 45,000 rpm in a TLA 45 rotor). Determination of RNase activity was done in a total volume of 200 µl containing 5 or 50 µl of supernatant of the second centrifugation step and 80 µg of Poly(rU)(Pharmacia) in RNase-assay buffer (40 mM Tris-acetate (pH 6.5), 0.5 mM EDTA, 5 mM dithiothreitol (DTT)). After incubation of the reaction mixture at 37°C for 1 hour 200 µl of 1.2 M perchloric acid, 20 mM LaSO₄ was added. After 15 min incubation on ice the mixture was centrifuged for 15 min at 4°C and 14,000 rpm in an Eppendorf centrifuge. To the supernatant 3 volumes of water were added and an aliquot of the mixture was analyzed by measuring the optical density at 260 nm using an Ultrospec 3000 spectrophotometer (Pharmacia). The mutation introduced into the E^{ms} gene completely abrogated RNase activity (Table 5).

To prove infection, the cells of the second dish were analyzed by immunofluorescence with BVDV specific monoclonal antibodies (Weiland et al., 1989) and found 100% positive.

Table 5

	CP7	B-349-d	control
OD ₂₆₀	2.5	1.1	1.1

Description of table 5

CP7: virus recovered from pA/BVDV/Ins-; B-349-d: virus recovered from pA/B-349-d; control; extract from noninfected MDBK cells.

Figure 1 Amino acid sequence of the glycoprotein E^{ns} of BVDV CP VII

5 1 ENITQWNLQDNGTEGIQRAMFQRGVNR**SLHGIWPEK**ICTGVPSHLATDTE
51 LKAIHGMMMDASEKTNYTCCRL**QRHEWNKHGWCN**WYNIEPWILLMNKTQAN
101 LTEGQPLRECAVTCRYDRDSDLNVVTQARDSPTPLTGCKKGKNFSFAGIL
10 VQGPCNFEIAVSDVLFKEHDCTSVIQDTAHYLVDGMTNSLESARQG TAKL
151
201 TTWLGRQLGILGKKLENKSKTWFGAYA

15 Sequences containing the residues conserved with regard to known RNases are underlined. The two histidine residues supposed to be in the catalytic centre of the enzyme are shown in bold characters. The shown sequence corresponds to residues 271 to 497 of the CP 7 polyprotein

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Claims:

1. Use of a live BVDV (bovine viral diarrhea virus), wherein the RNase activity residing in its glycoprotein E^{RNS} is inactivated for the preparation of a live vaccine for use in the prevention and/or treatment of BVDV infections in breeding stocks of cattle.
2. Use of a BVDV according to claim 1 for use in the prevention and/or treatment of BVDV infections in pregnant cows.
3. Use of a BVDV according to claim 1 for inducing fetal protection against BVDV infections in pregnant cows.
4. Use of a BVDV according to any one of claims 1 to 3 for use in the prevention and/or treatment of BVDV type I infections.
5. Use of a BVDV according to any one of claims 1 to 3 for use in the prevention and/or treatment of BVDV type II infections.
6. Use of a BVDV according to any one of claims 1 to 5, wherein said RNase activity is inactivated by deletions and/or mutations of at least one amino acid of said glycoprotein.
7. Use of a BVD virus according to any one of claims 1 to 6 comprising a BVDV pestivirus, wherein said RNase activity is inactivated by the deletion of the histidine residue at position 349, as described in figure 1 for the CP7 strain in an exemplary manner or corresponding thereto in other BVDV strains, of said glycoprotein.

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(54) Title: SAFE ATTENUATED BOVINE VIRAL DIARRHEA VIRUSES FOR USE IN PREGNANT COWS

(57) Abstract: This invention relates to the use of specifically attenuated live BVD (bovine viral diarrhea) viruses for the preparation of a vaccine for use in the prevention and/or treatment of BVDV infections in breeding stocks of cattle, pregnant cows and for fetal protection in pregnant cows.

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A. CLASSIFICATION OF SUBJECT MATTER

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C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
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A	V. VASSILEV ET AL.: "Authentic and full-length genomic cDNA clones of BVDV that yield infectious transcripts" J. VIROL., vol. 71, no. 1, 1997, pages 471-478, XP002144456 the whole document	1-7
A	WO 95 12682 A (THE UPJOHN COMPANY) 11 May 1995 (1995-05-11) page 13, line 16 - line 29	1-7
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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

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A	<p>M. HULST ET AL.: "Inactivation of the Rnase activity of glycoprotein Erns of classical swine fever virus results in a cytopathogenic virus" J. VIROL., vol. 72, no. 1, 1998, pages 151-157, XP002086212 the whole document</p>	1-7
A	<p>J. WINDISCH ET AL.: "Rnase of classical swine fever virus: biochemocal characterisation and inhibition by neutralising antibodies" J. VIROL., vol. 70, no. 1, 1996, pages 352-358, XP002095267 the whole document</p>	1-7

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